

Pooja Advani<sup>1</sup>, Sachin Kumar Deshmukh<sup>2</sup>, Sharon Wu<sup>2</sup>, Jacob Andring<sup>2</sup>, Joanne Xiu<sup>2</sup>, Jose P. Leone<sup>3</sup>, Priya Jayachandran<sup>4</sup>, Stephanie L. Graff<sup>5</sup>, Matthew Oberley<sup>2</sup>, George W. Sledge Jr<sup>2</sup>, Asher Chanan-Khan<sup>1</sup>

1. Division of Hematology and Oncology, Mayo Clinic, Jacksonville, FL, 2. Caris Life Sciences, Phoenix, AZ, 3. Department of Medicine, Dana-Farber Cancer Institute, Boston, MA, 4. Norris Comprehensive Cancer Center, Keck School of Medicine, University of Southern California, Los Angeles, CA, 5. Lifespan Cancer Institute, Legorreta Cancer Center, Brown University, Providence, RI

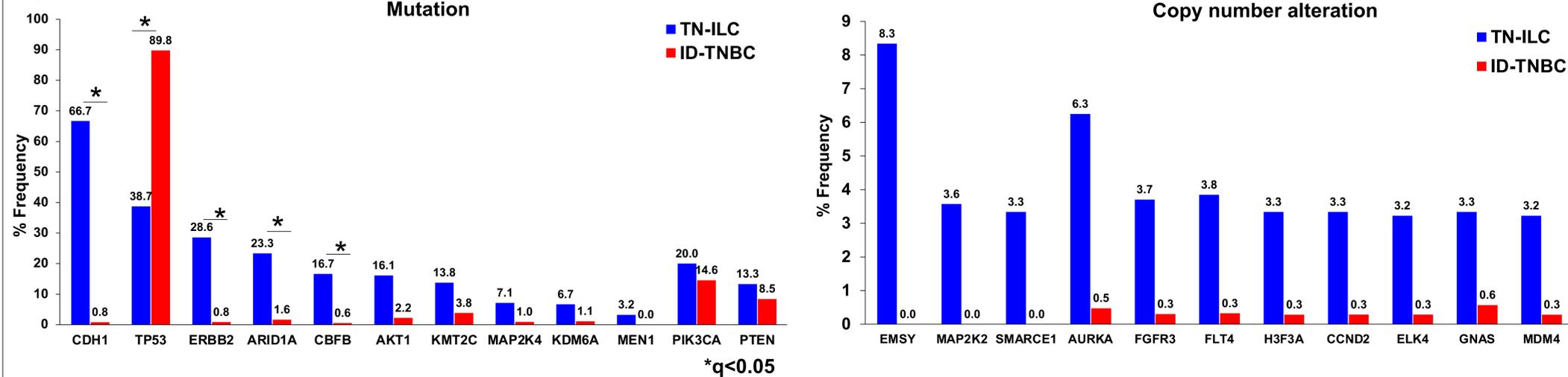
## BACKGROUND

Triple-negative invasive lobular carcinoma (TN-ILC) is a rare (0.1-1.4%) breast cancer with prognosis worse than ER positive ILC. Currently there are no targeted therapies or clinical trials specifically for TN-ILC. A comprehensive analysis of the molecular and immune landscape can help identify novel targets and pathways for TN-ILC to improve patient outcomes. Here, we characterized the molecular and immune signature of TN-ILC.

## METHODS

- 395 BC samples (Invasive ductal (ID) TNBC, n=364; TN-ILC, n=31) were analyzed by NGS (592, NextSeq; WES, NovaSeq), WTS (NovaSeq) (Caris Life Sciences, Phoenix, AZ).
- A total of 10 pro-apoptotic (*BAX*, *BAK1*, *BID*, *BAD*, *BIK*, *BCL2L11*, *BMF*, *HRK*, *PMAIP1*, *BBC3*) and 6 anti-apoptotic (*BCL2*, *BCL2L1*, *BCL2L2*, *MCL1*, *BCL2A1*, *BCL2L10*) BCL2 family genes were analyzed.
- Tumor mutational burden (TMB) totaled somatic mutations per tumor (high >10 mt/MB) was tested by IHC and NGS.
- Immune cell fractions were calculated by deconvolution of WTS: Quantiseq.
- Pathway enrichment was determined by GSEA (Broad Inst).
- Statistical significance was determined using chi-square and Mann-Whitney U test with p-values adjusted for multiple comparisons ( $q < 0.05$ ).

**Figure 1.** Mutation and copy number alteration analysis of TN-ILC and ID-TNBC



**Figure 2.** Immune cell infiltration in TN-ILC and ID-TNBC

Cells	TN-ILC	ID-TNBC
B cells	4.40	4.46
Mφ M1	2.17	3.46 *
Mφ M2	5.34	2.94 *
DC	3.00	3.21
Neutrophils	4.83	2.65 *
NK cells	3.15	3.05
T cells CD8	0.11	0.58
Tregs	1.51	1.99

■ Low Median% ■ High Median%  
 TN-ILC had higher infiltration of M2 macrophages and neutrophils but lower infiltration of M1 macrophages and CD8 T cells. Median% of monocytes and T cells CD4 were 0.0 in both the groups \* $q < 0.05$ .

**Figure 3.** Immune checkpoint gene expression

Gene	TN-ILC	ID-TNBC
CD274	2.78	4.01 *
PDCD1	0.24	0.52
CTLA4	0.82	1.61 *
PDCD1LG2	1.33	1.67
FOXP3	2.22	2.79
HAVCR2	14.98	19.70
LAG3	0.67	1.05 *
IDO1	1.07	2.90 *

■ Low Median ■ High Median  
 TN-ILC had decreased *CD274*, *CTLA4*, *FOXP3*, *LAG3* and *IDO1*. \* $q < 0.05$ .

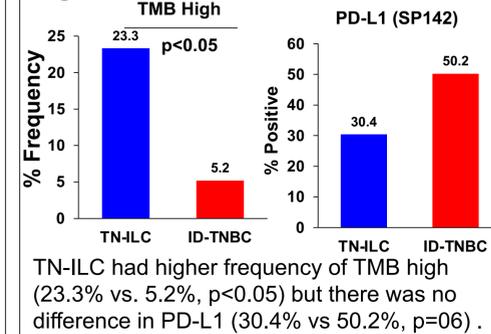
**Figure 4.** BCL2 family gene expression

Gene	TN-ILC	ID-TNBC
BAX	22.45	32.86 *
BAK1	3.85	5.16 *
BID	9.57	15.47 *
BAD	10.59	8.59
BIK	3.24	1.46 *
BCL2L11	18.11	22.43
BMF	6.73	7.61
HRK	0.00	0.11 *
PMAIP1	0.27	1.70 *
BBC3	0.92	0.83
BCL2	2.06	2.25
BCL2L1	18.39	18.41
BCL2L2	2.48	2.30
MCL1	17.08	25.86 *
BCL2A1	2.22	8.42 *
BCL2L10	0.20	0.38 *

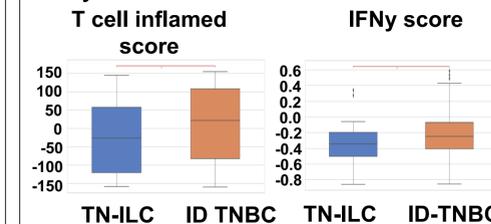
■ Low Median ■ High Median  
 TN-ILC had differential BCL2 family gene expression. \* $q < 0.05$ .

## RESULTS

**Figure 5.** Immune marker analysis

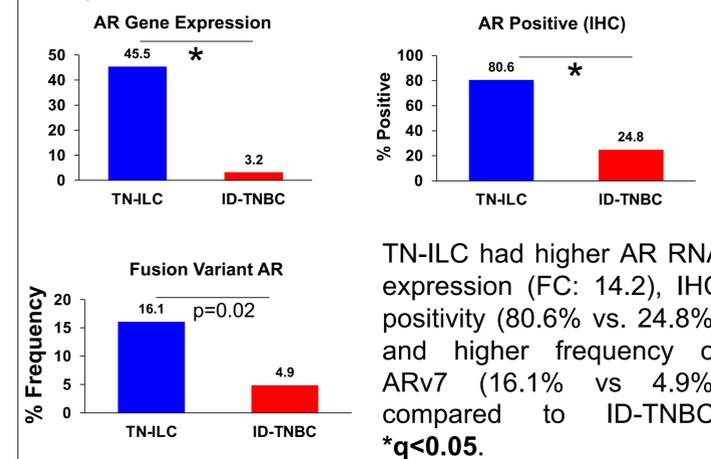


**Figure 6.** T cell inflamed and IFNγ score

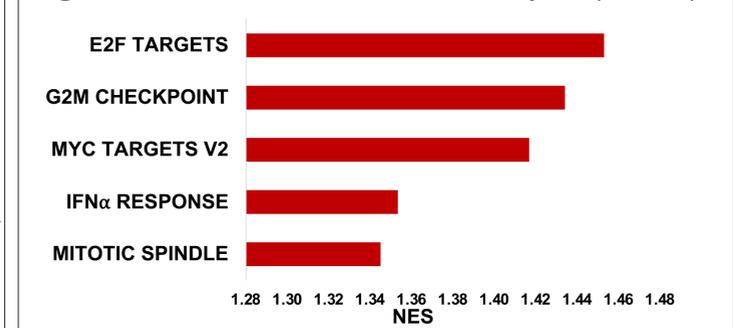


TN-ILC had decreased T cell inflamed score (-0.27 vs 21.5) and IFNγ score (-0.34 vs -0.24, all  $q < 0.05$ ).

**Figure 7.** AR expression and fusion variant AR analysis



**Figure 8.** Gene set enrichment analysis (GSEA)



ID TNBC had gene set enrichment of the E2F targets, G2M checkpoint, MYC targets V2, IFNα response and mitotic spindle pathways compared to TN-ILC (all  $FDR < 0.25$ ).

## CONCLUSIONS

These data suggest that TN-ILC had higher frequency of *CDH1*, *ERBB2*, *AKT1*, *ARID1A* mutations, higher M2 macrophages and neutrophils and lower M1 macrophages and CD8 T cells infiltration and, lower T cell inflamed signature. High TMB and AR expression can translate into use of immunotherapy (ICI) and AR antagonists in these patients.