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# Background

- The main mismatch-binding factor in humans is hMutSa, consisting of MSH2 and MSH6, which recognizes single-base mispairs. Upon mismatch binding, the hMutS complex undergoes conformational change into a sliding clamp and a hMutL heterodimer is recruited.
- hMutL complex is hMutLa, consisting of MLH1 and PMS2 and • The main participating in the repair of single-base mismatches. When the hMutS-hMutL complex encounters a strand discontinuity, an excision machinery is recruited, the mismatch containing fragment is degraded, and a new strand synthesized

# Figure 1: The different hMutS and hMutL complexes in human MMR.



- Salem et al. reported that in CRC and EC, loss of co-expression of MLH1/PMS2 was more common than loss of MSH2/MSH6 (P < .0001). Loss of coexpression of MLH1/PMS2 was associated with lower mean TMB (MLH1/PMS2: 25.03 mut/Mb vs MSH2/MSH6 46.83 mut/Mb; P < .0001).
- In colorectal cancer (CRC) and endometrial cancer (EC) patients (pts), preliminary data suggest a differential response to immune checkpoint inhibitors (ICIs) according to different MMR alterations.
- The drivers of this difference remain unknown and no reliable predictive biomarker has been found.
- We explored the genomic alterations, tumor mutation burden (TMB), immune-related gene expressions and signatures, tumor microenvironment (TME), neoantigen load and median overall survival (mOS) in CRC and EC pts treated with ICIs with different MMR alterations.

# **Materials and Methods**

- CRC (N= 14,949) and EC (N=3,574) specimens were tested at Caris Life Sciences (Phoenix, AZ) with Next Gen Sequencing (NGS) of DNA (592-gene or whole exome sequencing) and RNA (whole transcriptome sequencing).
- MMR/MSI status was determined by IHC of MMR protein and/or NGS.
- Immune cell abundance was quantified using quanTIseq.
- Gene expression profiles were analyzed for T cell-inflamed signature (TIS) and IFN-gamma scores.
- Immune epitope prediction was performed using the NetMHCpan v4.0 method in the Immune Epitope Database.
- Real-world mOS was obtained from insurance claims data and calculated from tissue collection or ICIs start to last contact.
- Statistical significance was determined using chi-square/Fisher-Exact and adjusted for multiple comparisons (adjusted p < 0.05).

# The differential response to immune checkpoint inhibitors in colorectal and endometrial cancer patients according to different mismatch repair alterations

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# Figure 2: Colorectal (CRC) and Endometrial (EC) Patients

### **Colorectal Cancer** (N = 14949) neg MSH2 MSH2 pos MSH6 MSH6 MSH6 MSH6 MSH2

84 (0.6%) patients had MutS co-loss 648 (4.7%) patients had MutL co-loss 117 (0.9%) patients had other MMR IHC loss.

MSH6

MSH6

### pos neg MSH2 MSH2

Endometrial Cancer (N = 3574)

PMS2	pos	pos		neg		pos		neg	
		MSH6		MSH6		MSH6		MSH6	
		pos	neg	pos	neg	pos	neg	pos	neg
		2271	28	2	48	1	0	0	0
	neg	MSH2				MSH2			
		pos		neg		pos		neg	
		MSH6		MSH6		MSH6		MSH6	
		pos	neg	pos	neg	pos	neg	pos	neg
		44	3	0	0	915	2	0	1

48 (1.4%) patients had MutS co-loss 915 (27.6%) patients had MutL co-loss 81 (2.4%) patients had other MMR IHC loss.

# **Figure 3: Genomic alterations**



MSH2, TP53, ERBB3, PPP2R1A, IHC-PD-L1, MSH3, ERBB2 and ARHGAP35 positive rates are significantly higher in MutS.

IHC PR and IHC ER loss positive rates are significantly lower in MutS

# Figure 4: Tumor Mutation Burden



# **Colorectal Cancer**

# **Endometrial Cancer** MLH1 and PMS2 co-loss and MSH2 and MSH6 co-loss normal MSH2 and MSH6 and normal MLH1 and PMS2

TMB values are significantly higher in MutS co-loss

# Figure 5: Immune related gene expression

# **Colorectal Cancer**

Expression of CD274, IFNG, IDO1 and LAG3 are significantly lower in MutS coloss

## **Endometrial Cancer**

Expression of CD80, CD274, CTLA4, HAVCR2, IFNG, IDO1, LAG3, PDCD1 and PDCD1LG2 are significantly higher in MutS co-loss

### **Colorectal Cancer**

pos

MSH6

pos | neg | pos | neg | pos | neg

MSH6



Mutation frequencies of APC, KRAS, ERBB2, ERBB3 are significantly higher in MutS. IHC PD-L1 positive rate, mutation frequency of BRAF is lower in MutS

# **Endometrial Cancer**

# Results

# Figure 6: Immune signature (T-cell inflamed score)



No difference in T-cell inflamed score betweer

the two groups



**Endometrial Cancer** 

T-cell inflamed score is significantly higher in MutS co-loss

# Figure 7: Immune signature (IFN-gamma score)





INF-gamma score is significantly higher in MutS co-loss

# **Figure 8: Tumor Microenvironment**

### **Colorectal Cancer**

IFN-gamma scores are significantly lower in

MutS co-loss group.

Cell Type	fold change	
B cell	0.9299854	
Macrophage M1	1.0353868	
Macrophage M2	0.7504589	
Monocyte	inf	
Neutrophil	1.0647099	
NK cell	0.8898786	
T cell CD4+ (non-		
regulatory)	1.0405909	
T cell CD8+	1.4589834	
T cell regulatory		
(Tregs)	1.0091382	
Myeloid dendritic		
cell	0.97253186	

### B cells, Macrophage M2 (cold) and NK cells are significantly higher in MutS co-loss

# **Endometrial Cancer**

Cell Type	fold change
B cell	1.0926403
Macrophage M1	0.61074114
Macrophage M2	1.1284298
Monocyte	3.4750133
Neutrophil	1.0382272
NK cell	1.0406753
T cell CD4+ (non-	
regulatory)	0.8124716
T cell CD8+	0.32283562
T cell regulatory	
(Tregs)	0.8363445
Myeloid dendritic	
cell	1.2676436

Macrophage M1 and CD8+ cell are significantly higher in MutS co-loss

Myeloid dendritic cell is significantly lower in MutS

# Figure 9: Neoantigen load (number of neoepitopes)



The numbers of high, intermediate, and low affinity neoepitopes are significantly higher in MutS co-loss

**Endometrial Cancer** 8888

The numbers of high, intermediate, low affinity epitopes are significantly higher in MutS co-loss



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# Figure 10: Median Overall Survival (collection to last contact)

# **Colorectal Cancer**



<sup>149)</sup> vs. MutL co-loss (N = 980) was 56 months (m) vs. 36 m (p = 0.003)

## **Endometrial Cancer**



loss (N = 1804) was NR vs. 47 m (p < 0.001)

# Figure 11: Median Overall Survival (ICIs treatment to last contact)



- In ICI-treated pts, the mOS in MutS co-loss (N = 28) vs. MutL co-loss (N = 149) was not reached (NR) vs. 32 m (p = 0.005).
- BRAF mutation didn't impact survival in MutL co-loss

## **Endometrial Cancer**



In ICI-treated pts, the mOS in MutS co-loss (N = 11) vs. MutL co-loss (N = 273) was NR vs. NR (p = 0.559)

# Summary and Conclusion

Features	Colorectal Cancer (MutS co-loss vs. MutL co-loss)	Endometrial Cancer (MutS co-loss vs. MutL co-loss)
Incidence	0.6 vs 4.7	1.4 vs 27.6
Genomic alterations	High APC, KRAS, ERBB2, ERBB3 vs. PD-L1 and BRAF	MSH2, TP53, ERBB3, PPP2R1A, IHC-PD-L1, MSH3, ERBB2 and ARHGAP35 vs. ER and PR
ТМВ	Higher in <u>MutS</u> co-loss	Higher in MutS co-loss
T-cell inflamed score	No difference	Higher in MutS co-loss
IFN-gamma score	Lower in MutS co-loss	Higher in MutS co-loss
Immune related gene expression	CD274, IFNG, IDO1 and LAG3 are lower in MutS co-loss	CD80, CD274, CTLA4, HAVCR2, IFNG, IDO1, LAG3, PDCD1, and PDCD1LG2 are higher in MutS co-loss
Tumor microenvironment	B cells, Macrophage M2 (cold) and NK cells are higher in MutS co-loss.	Macrophages M1 and CD8+ cell are higher in MutS. Myeloid dendritic cell is lower in MutS co-loss.
Neoantigen Load	High, intermediate, and low affinity neoepitopes are higher in <u>MutS</u> co-loss	High, intermediate, and low affinity neoepitopes are higher in <u>MutS</u> co-loss
mOS (collection to last contact)	56 months (m) vs. 36 m (p = 0.003).	NR vs. 47 m (p < 0.001)
mOS( ICIs treatment to last contact)	NR vs. 32 m (p = 0.005)	NR vs. NR (p = 0.559)

- This is the largest study to explore differential response to ICIs in CRC and EC pts with different MMR alterations.
- In pts with CRC and EC, the mOS was longer in MutS co-loss compared to MutL co-loss.
- In ICI-treated pts, the mOS was longer in MutS co-loss compared to MutL co-loss in CRC but not in EC.
- Apart from TMB, among the explored biomarkers, neoantigen load was higher in MutS co-loss compared to MutL co-loss in both CRC and EC and maybe the driving factor for differential response to ICIs.